

Extracellular ATP and P2X7 receptors in neurodegeneration

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Abstract

Neuronal injury and cell death in the central nervous system (CNS) are underlying features of neurodegenerative disorders. However, our understanding of the fundamental mechanisms involved is still limited. Inflammatory processes mediated by cytokines, and interleukin-1 (IL-1) in particular, play a significant role in neuronal death following pathological insults. Despite this growing area of research, very little is known about the factors regulating the expression, cleavage and release of interleukin-1 in the brain. Recent studies on immune cells demonstrate that extracellular ATP can act as a potent stimulus for the maturation and release of interleukin-1 β , via activation of P2X7 receptors. Stimulation of P2X7 receptors with ATP has dramatic cytotoxic properties and a wider role in neurodegenerative processes is possible. This review discusses the potential involvement of extracellular ATP and P2X7 receptors as regulators of interleukin-1-mediated neuropathologies and thus as a mediator of cell death following pathological insults.

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1. Introduction—ATP and neurodegeneration

The role of extracellular ATP and purinoceptors in cytokine regulation and neurodegeneration is the focus of a rapidly expanding area of research. ATP can act as a neurotransmitter, whilst the presence of P2X7 receptors (purinergic receptor sub-class) on immune cells suggests that it also regulates immune function and inflammatory responses. In addition, activation of this receptor has dramatic cytotoxic properties, which together with its ability to regulate cytokine production and release, suggest that it can act as an important regulator of cell death in response to pathological insults.

Neurodegeneration is the underlying basis of many disorders including cerebral ischaemia, brain trauma, multiple sclerosis, Parkinson's, Alzheimer's and Huntington's diseases. Despite being the focus of considerable research over the past 20 years, our understanding of the fundamental mechanisms involved in neurodegeneration is still limited in terms of potential clinical therapies. This review will discuss some recent progress in the field of neurodegeneration, the involvement of inflammatory cytokines such as interleukin-

1 (IL-1), and will elaborate new findings on their regulation through ATP stimulation of P2X7 receptors, which may offer novel therapeutic approaches.

2. Mechanisms of neurodegeneration

The mechanisms involved in neuronal death are complex and are likely dependent upon multiple signalling cascades. This review will focus on cerebral ischaemia, as a major clinical condition for which there is extensive experimental data. However, a diverse range of neurodegenerative diseases, trauma and excitotoxicity show common features. Cerebral ischaemia occurs when the blood supply to an area of the brain is reduced either permanently or transiently, resulting in oxidative and metabolic stress to the tissue. One consequence of such metabolic compromise is the depletion of cellular ATP reserves. With the failure of the Na⁺/K⁺ pump, neurones depolarise resulting in excess release and accumulation of the excitatory amino acid neurotransmitter, L-glutamate (Choi, 1994; Haddad and Jiang, 1992; Limbrick et al., 1995). Glutamate activates a number of post-synaptic ionotropic (e.g., NMDA, kainate and AMPA) and metabotropic receptors leading to disturbances in the intracellular ionic environment. High cytoplasmic Ca²⁺ concentrations, resulting from suppression of Ca²⁺ pumps and activation of

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agonist-dependent and voltage-gated Ca^{2+} channels, are central to the pathological process culminating in cell death (Siesjö, 1981).

Neurones, which survive the primary insult, may die hours or days later as increased intracellular Ca^{2+} or other second messenger systems initiate cascades with the activation of proteases, lipases, nitric oxide synthases and nucleases. Depending on the severity of the insult and the metabolic energy state of the cell, death may occur by either apoptotic or necrotic mechanisms (Lee et al., 1999; Nicotera, 2000). Increased apoptosis is reported in several neurodegenerative diseases (Forloni et al., 1996; Saudou et al., 1998; Kim et al., 1999). In experimentally induced cerebral ischaemia in rodents, apoptosis may be particularly important in the penumbra. This area, suffers only mild ischaemic insult, but may undergo apoptosis from actions of secondary mediators such as oxygen radicals, cytokines and lipid peroxidation products from the more severely stressed areas of the necrotic core (Mattson, 1998; Charriaut-Marlangue et al., 1996; Lipton and Rossenber, 1994). During reperfusion of the tissue, excessive uptake of Ca^{2+} by mitochondria could play a key role in ischaemic cell death by activating the production of oxygen-free radicals and blocking the synthesis of high-energy phosphates (Kristian and Siesjö, 1998).

Alterations in Ca^{2+} homeostasis result in changes in protein synthesis and secretion, which involve the activation of nuclear factor κB (NF- κB) and hence, NF- κB -dependent genes, such as interferons and cytokines, and activation of death related genes that increase caspase expression (Paschen and Douthell, 1999; Nicholson and Thornberry, 1997). Multiple caspases are activated during ischaemia (Bhat et al., 1996; Chen et al., 1997; Harrison et al., 2001). Pan-caspase inhibitors are neuroprotective in a number of experimental neurodegenerative paradigms (Schneider et al., 2000), whilst ischaemic lesions are significantly reduced in mice lacking the caspase-1 gene (Li et al., 1995; Liu et al., 1999).

For more than 15 years, extracellular glutamate accumulation has been suspected as the initiator of immediate and delayed neuronal death following stroke (McCulloch et al., 1993). Despite this, inhibition of excess glutamate by antagonism of receptors, or the enhancement of glutamate re-uptake mechanisms have not provided effective stroke therapies (Doble, 1999). Since glutamate transmission, Ca^{2+} homeostasis and nitric oxide (NO) synthesis are all involved in normal brain function, therapeutic strategies aimed at these areas may be accompanied by unacceptable side effects (Iadecola, 1997; Doble, 1999).

3. Neurodegeneration and cytokine regulation

Evidence produced over the last decade has demonstrated an important role for inflammatory processes in the pathogenesis of cerebral ischaemia and many forms of neurodegeneration (Kogure et al., 1996; Allan and Rothwell, 2001). Cytokines are a group of polypeptides, which include

interleukins, tumour necrosis factors, chemokines, interferons and growth factors/neurotrophins, and are key mediators of inflammation. Members of each of these polypeptide families have been implicated in stroke either as mediators of damage or potential neuroprotective/neurotrophic factors. Since cytokines appear to play little or no role in normal brain function therapeutic approaches to block their actions may prove successful. Indeed, the involvement of cytokines in central nervous system (CNS) disease, is now a rapidly growing area of biological and clinical research (Barone and Feuerstein, 1999; del Zoppo et al., 2000). An early ischaemic event is the infiltration of leukocytes and the activation of resident brain cells including astrocytes, microglia and endothelial cells (Stoll et al., 1998). This inflammatory reaction is triggered by expression of a complex array of genes regulated by NF- κB , including the inflammatory cytokines (interleukin-1 β (IL-1 β), tumour necrosis factor- α (TNF- α) and interleukin-6 (IL-6), and inducible nitric oxide synthase (iNOS) (see Feuerstein et al., 1998 for review)), which modulate inflammation, apoptosis and neuronal plasticity.

Cytokine bioactivity can be regulated at the levels of transcription, translation, cleavage and cellular release as well as through receptor and post-receptor signalling mechanisms. Excitatory amino acids can regulate cytokine expression directly during excessive release following CNS injury, through phosphorylation of C-jun N-terminal kinase (JNK) (Sattler et al., 1999; Savinainen et al., 2001) and the activation of the transcriptional factor *c-jun* which promotes expression of cytokines (e.g., IL-1, IL-6 and TNF).

The cytokine interleukin-1 has received the most attention as a potential mediator of neurodegeneration. The interleukin-1 family comprises two agonists, interleukin-1 α and -1 β , and a naturally occurring interleukin-1 receptor antagonist (IL-1ra) (Dinarello, 1998; Fantuzzi and Dinarello, 1999; Hannum et al., 2002). Interleukin-1's actions have so far been ascribed to binding to the interleukin-1 receptor 1 (IL-1R1, 80 kDa) which then interacts with a specific accessory protein (AcP) to recruit intracellular adapter molecules and signal through mitogen-activated protein (MAP) kinases and NF- κB (Sims et al., 1993; O'Neill and Dinarello, 2000). However, the possibility of other ligands and receptors exists (Smith et al., 2000; Touzani et al., 2002).

Interleukin-1 α , interleukin-1 β , interleukin-1 receptor antagonist and the enzyme responsible for interleukin-1 β maturation, caspase-1 (originally termed interleukin-1 converting enzyme—ICE), are all up regulated rapidly in response to experimentally induced ischaemic brain damage in rodents (Loddick et al., 1997; Davies et al., 1999; Legos et al., 2000; Touzani et al., 2002) and raised interleukin-1 and interleukin-1 receptor antagonist levels have been reported in human stroke patients (see Touzani et al., 1999). Administration of interleukin-1 to the brain of normal rodents or to neurones in culture is not cytotoxic, but in vivo administration of low doses of interleukin-1 (i.c.v.) exacerbates ischaemic and excitotoxic damage in rat

and mouse models (Lawrence et al., 1998; Stroemer and Rothwell, 1998; Loddick and Rothwell, 1996; Yamasaki et al., 1995). A number of studies have now demonstrated that inhibiting the release or actions of interleukin-1 markedly reduces damage in permanent focal ischaemia in the rodent (Relton and Rothwell, 1992; Garcia et al., 1995; Touzani et al., 2002). Furthermore, ischaemic damage is reduced by over 70% in mice lacking genes for both interleukin-1 α and -1 β (Boutin et al., 2001). Targeting interleukin-1 could thus offer therapeutic strategies, and interleukin-1 receptor antagonist is now being tested in phase II clinical trials.

Interleukin-1 is a key mediator of host responses, inflammation and tissue injury, and is produced abundantly by macrophages and monocytes. In the brain, Interleukin-1 β is expressed and released mainly by microglia, although astrocytes and invading macrophages may also contribute to interleukin-1 β production some time after the insult (Pearson et al., 1999; Mabuchi et al., 2000). Microglia are the phagocytic inflammatory mediators within the CNS and they respond to diverse insults by adopting an ameboid morphology, proliferating and producing reactive oxygen species as well as a number of inflammatory mediators such as interleukin-1 and TNF α (Kreutzberg, 1996; Raivich et al., 1999).

Interleukin-1 β is expressed as an inactive precursor (pro IL-1 β) which is increased by pro-inflammatory stimuli such as bacterial endotoxin (lipopolysaccharide—LPS; (Dinarello, 1991)), other inflammatory cytokines (including interferon- γ (IFN γ) and TNF α (Dinarello and Krueger, 1986; Roux-Lombard, 1998), or activated T-lymphocytes (Vey et al., 1997). In vitro, the cleavage of pro-interleukin-1 β and release of the mature active cytokine in lipopolysaccharide-stimulated macrophages, monocytes or microglia, requires a secondary stimulus (e.g., ATP) for the activation of caspase-1 (Hogquist et al., 1991; Thornberry et al., 1992; Cerretti et al., 1992; Sanz and Di Virgilio, 2000) and subsequent release of the mature cytokine.

4. ATP and the regulation of IL-1

In addition to its key roles as a ubiquitous enzyme cofactor and a source of cellular energy, the purine nucleotide ATP also functions as a potent extracellular messenger and neurotransmitter via activation of members of the P2 receptor family (Ralevic and Burnstock, 1998). P2 receptors comprise both metabotropic P2Y and ionotropic P2X receptor families. Seven members of the P2X receptor family have been described, although a P2X8 receptor has been suggested (Ralevic and Burnstock, 1998; Bo et al., 2000).

4.1. P2X7 receptors

Since their discovery in immune cells (Hogquist et al., 1991), P2X7 receptors have been proposed as mediators of

inflammation, and a potential role in neurodegeneration has been suggested (Di Virgilio, 1995). The P2X7 receptor (previously referred to as the cytolytic P2Z receptor) has been implicated in signalling between macrophages and other immune target cells (Surprenant et al., 1996). The P2X7 receptor shares 35–40% homology with other P2X receptors. It has two hydrophobic membrane-spanning domains and an extracellular loop, and forms transmembrane ion channels. Stimulation of the P2X7 receptor with high concentrations of ATP (its major endogenous ligand) triggers massive transmembrane ion fluxes (particularly influx of Ca²⁺ and Na⁺, and efflux of K⁺) and the formation of non-selective plasma membrane pores (Ras-sendren et al., 1997) that result in cell death (Di Virgilio et al., 1998a). In addition, P2X7 receptor activation stimulates the induction of multiple cytokine pathways that may co-ordinate inflammatory responses.

Under normal conditions, extracellular nucleotides are present in only low concentrations. However, activated immune cells (lymphocytes (Filippini et al., 1990), macrophages (Sikora et al., 1999), microglia (Ferrari et al., 1997a), and platelets (Beigi et al., 1999)), and dying cells may release high concentrations of different nucleotide di- and tri-phosphates into the extracellular space (Dubyak and el-Moatassim, 1993). Extracellular ATP concentrations increase significantly under inflammatory conditions in vivo (Lazarowski et al., 2000; Di Virgilio et al., 1998b), and in response to tissue trauma (e.g., ischaemia/hypoxia; Nieber et al., 1999). ATP can be co-released with glutamate from pre-synaptic nerve terminals (Sperlágh and Vizi, 1996).

4.2. Interleukin-1 production

ATP was first shown to stimulate the release and processing of interleukin-1 β and induce cell death in murine macrophages by Hogquist et al. (1991). ATP induces the processing and release of interleukin-1 β by activation of P2X7 receptors (Surprenant et al., 1996; Ferrari et al., 1997b; Perregaux and Gabel, 1998; Solle et al., 2001) and cytokine release can be prevented by inhibition of caspase-1 (Watanabe et al., 1998). Caspase-1 is constitutively expressed in most cells as a 45-kDa inactive precursor, which requires two cleavages to form an active heterodimer (Wilson et al., 1994; Gu et al., 1995). The mechanism of caspase-1 activation was first suggested to be autoproteolysis (Thornberry et al., 1992). More recent evidence suggests that P2X7 receptor stimulation activates caspase-1 through the depletion of intracellular K⁺ (Perregaux and Gabel, 1994; Schulze-Lohoff et al., 1998; Ferrari et al., 1999a; Laliberte et al., 1999; Humphreys et al., 2000). Priming of cells with lipopolysaccharide renders cells particularly sensitive to the cytotoxic and cytokine releasing actions of ATP, possibly through the increased expression of pro-caspase-1 (Le Feuvre et al., 2002). ATP may also affect interleukin-1 β production at transcriptional and translational levels, since P2X7 depend-

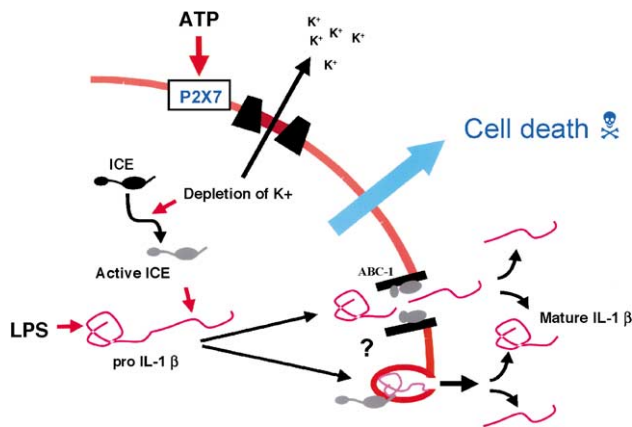


Fig. 1. Scheme depicting proposed mechanisms of ATP stimulated IL-1 β processing and release.

ent induction of the transcriptional activator, NF- κ B, has been observed in microglia (Ferrari et al., 1997c; Humphreys et al., 2000) (Fig. 1).

4.3. IL-1 release

The mechanism underlying cellular release of mature interleukin-1 β in response to ATP and indeed other stimuli is unknown. Pro-interleukin-1 β is synthesized on free ribosomes, lacks a signal sequence and therefore accumulates in the cytosol of activated cells. A small fraction of pro-interleukin-1 β is myristylated (Stevenson et al., 1993) and monocytes secrete small amounts of pro-interleukin-1 β by a mechanism which appears to be distinct from the secretion of the mature form (Beuscher et al., 1990). Interleukin-1 β release induced by ATP in lipopolysaccharide primed cells in vitro has proved a useful tool for studying these processes. ATP-induced release and maturation of interleukin-1 β from monocytes and macrophages is suppressed by inhibition of the anion exchanger function of the ATP-binding cassette-1 (ABC-1) (e.g., by sulfonylurea glyburide or ethacrynic acid). This suggests that anion conductance is a necessary component of ATP promoted externalisation of interleukin-1 (Laliberte et al., 1994; Perregaux et al., 1996; Hamon et al., 1997; Sanz et al., 1998). These inhibitors do not affect the K⁺ efflux, and thus caspase-1 activation would still occur (Perregaux and Gabel, 1998).

Bi-phasic changes in Ca²⁺ levels are observed in response to stimulation of P2X7 receptor expressing dendritic cells, with a rapid release of Ca²⁺ from intracellular stores and a delayed influx across the cell membrane (Ferrari et al., 2000). Current literature suggests that interleukin-1 β release and processing is Ca²⁺-independent but depends upon efflux of K⁺ (Perregaux et al., 1992; Perregaux and Gabel, 1994; Walev et al., 1995). Studies with Ca²⁺ ionophores revealed that Ca²⁺ influx has no effect on the release of mature interleukin-1 β , whilst agents which

induce K⁺ efflux, such as nigericin, stimulate interleukin-1 maturation and release. More recently K⁺ efflux was shown to regulate interleukin-1 β processing via the Ca²⁺-independent phospholipase A₂ (Walev et al., 2000). Other studies have reported conflicting results. Ciesynski (1999) suggested that a Ca²⁺ ionophore augments interleukin-1 secretion by lipopolysaccharide in chick macrophages, and West et al. (1996) found that lipopolysaccharide-stimulated interleukin-1 release does not require Ca²⁺-dependent signalling pathways.

Our recent research has demonstrated that ATP-induced IL-1 β processing and release requires not only K⁺ efflux, but also release of Ca²⁺ from intracellular stores (Brough et al., unpublished observations). The processing and release of IL-1 β in macrophages and microglia is dramatically inhibited by addition of the intracellular Ca²⁺ chelator [1,2-bis(*o*-Aminophenoxyethan-*N,N,N',N'*-tetraacetic acid tetra (acetoxymethyl) ester] (BAPTA-AM) (Fig. 2). Further

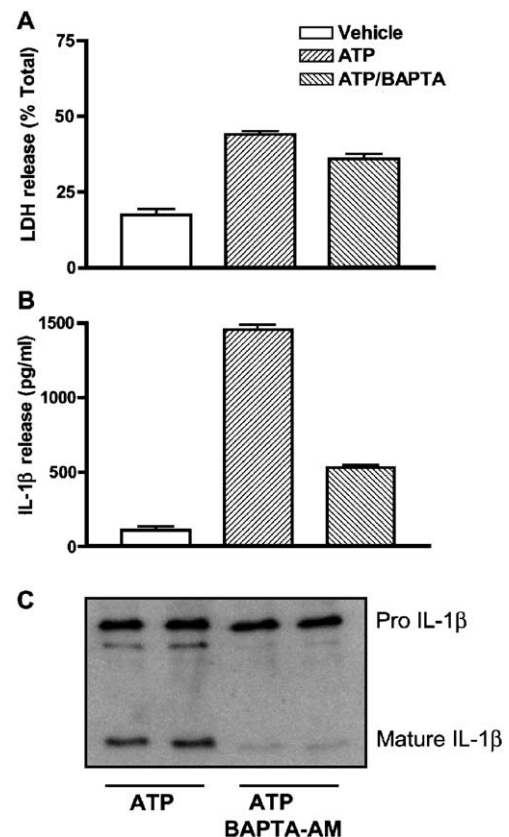


Fig. 2. The calcium chelator, BAPTA-AM, inhibits ATP induced interleukin-1 release in pure microglial cultures without affecting cell death. Primary microglia were primed with lipopolysaccharide (LPS, 0.1 μ g) overnight then treated with 5 mM ATP in the presence of vehicle or BAPTA-AM (30 μ M). (A) Cell death was assessed as lactate dehydrogenase (LDH) release and expressed as a % of total LDH (measured by complete cell lysis with Triton-X 100). (B) The concentration of IL-1 β released into the medium was measured by a specific murine IL-1 β ELISA (kindly supplied by Dr. S. Poole). (C) The presence of pro- and mature forms of IL-1 β was tested using Western blot analysis.

experiments have revealed that it is the release of Ca^{2+} from intracellular stores rather than an influx of extracellular Ca^{2+} that is involved. BAPTA-AM did not affect caspase-1 activity since caspase-1-induced cell death was still apparent (Le Feuvre et al., 2002).

Although the mechanism of mature interleukin-1 β release from the cell is not yet understood, there are a number of proposals resulting from work in monocytes. These currently suggest interleukin-1 release to depend upon a transmembrane pore gated by caspase-1 (Singer et al., 1995), secretion via late endosomes (Andrei et al., 1999) or secretion via microvesicles shed from the plasma membrane (MacKenzie et al., 2001). In contrast with our latest results, showing the importance of Ca^{2+} release from intracellular stores, the shedding of microvesicles in a human monocytic cell line (THP-1, MacKenzie et al., 2001) was dependent upon extracellular Ca^{2+} .

5. Cytotoxic effects of P2X7-receptor stimulation

Activation of P2X7 receptors is associated with cytotoxicity, and this receptor has been described as a death receptor (Di Virgilio et al., 1998a). It was initially suggested that the high concentrations of extracellular ATP required to stimulate cytotoxic actions of this receptor might not be reached in vivo. However, there is evidence that even short periods of P2X7 receptor activation are cytotoxic and that once activated, the P2X7 receptor sets in motion an irreversible death process (Hogquist et al., 1991; Laliberte et al., 1994). Cells primed with inflammatory mediators (e.g., lipopolysaccharide) are particularly susceptible to the toxic actions of ATP (Mehta et al., 2001; Le Feuvre et al., 2002) and this priming effect may alter the distribution or activation of P2X7 receptors in cell membranes (Denlinger et al., 2001).

We have demonstrated that the pan-caspase inhibitor z-Val-Ala-DL-Asp (OMe)-fluoromethylketone (zVAD-fmk) markedly inhibits cell death of macrophages exposed to ATP and LPS, and ATP fails to induce cell death of LPS primed macrophages from caspase-1 knock-out (KO) mice (Le Feuvre et al., 2002). These cytotoxic actions are independent of cytokine release (interleukin-1 α , - β or interleukin-18), suggesting that caspase-1 has additional actions possibly through induction of other caspases involved in cell death. Indeed, stimulation of P2X7 receptors leads to activation of multiple caspases (e.g., caspases 3 and 9 (Ferrari et al., 1999a)). Prolonged exposure to ATP may reflect induction of other death pathways, such as the activation of stress activated protein kinases (Humphreys et al., 2000). Most importantly, P2X7 regulation of multiple caspases may act as a switch between death by necrosis or apoptosis (Ferrari et al., 1999a), which could affect pathological outcome.

The transcriptional activator NF- κ B has been implicated in the control of apoptosis and pro-inflammatory gene

expression. A broad range of stimuli can activate NF- κ B (e.g., lipopolysaccharide and inflammatory cytokines). In particular, ATP can lead to selective activation of NF- κ B in microglial cells, through P2X7 stimulation and induction of caspase-1-like proteolytic activity (Ferrari et al., 1997c, 1999b). P2X7 receptors may also be involved in endotoxin signal transduction in macrophages (Beigi and Dubyak, 2000), since P2X7 receptor antagonists such as oxidised-ATP, attenuate a subset of endotoxin-induced effects including NF- κ B activation and up-regulation of inducible NO synthase. Thus, an association between endotoxic events and P2X7 receptors appears to exist.

Finally, P2X7 receptors may affect immune function through antigen presentation (Mutini et al., 1999), nitric oxide production (Molloy et al., 1994), cell–cell communication (Inoue, 1998), and the formation of gap junctions and multinucleated giant cells (Chiozzi et al., 1996). P2X7 receptors have been identified at pre-synaptic nerve terminals, suggesting a role in synaptic transmission (Deuchars et al., 2001) where ATP may affect glutamate release (Nakatsuka and Gu, 2001) and vice versa. Our observations that ATP induces alterations in Ca^{2+} homeostasis which affect interleukin-1 β processing and release, suggest that it could have a wider role in neurodegenerative processes.

6. Role of P2X7 in vivo

Involvement of P2X7 receptors in neurodegenerative conditions remains speculative since the role of the P2X7 receptor in vivo remains to be studied in detail. The only published reports so far have demonstrated that ATP increases release and processing of IL-1 β in the peritoneum following LPS injections through a P2X7-receptor dependent mechanism (Griffiths et al., 1995; Solle et al., 2001). High concentrations of extracellular ATP have been observed at sites of tissue trauma (Nieber et al., 1999), and increased microglial P2X7 receptor immunoreactivity has been described in the penumbra following cerebral ischaemia in rodents (Collo et al., 1997). Whether this reflects an increase in P2X7 receptor density per se, or is due to infiltration of microglia to the site of inflammation has not been clarified. We are currently investigating a potential role for the P2X7 receptor in neurodegeneration in vivo, looking specifically at its role in mediating cell death following cerebral ischaemia and excitotoxic lesions induced by striatal infusion of the NMDA receptor agonist methano-glutamate (M-Glu). Owing to the lack of selective P2X7 antagonists and agonists, initial studies have been conducted comparing normal mice and those in which the gene for the P2X7 receptor has been deleted (P2X7 knock-out mice). The P2X7 knockout mice exhibited “normal” responses to reperfusion and permanent ischaemia and to excitotoxic damage compared to their wild type counterparts (Fig. 3).

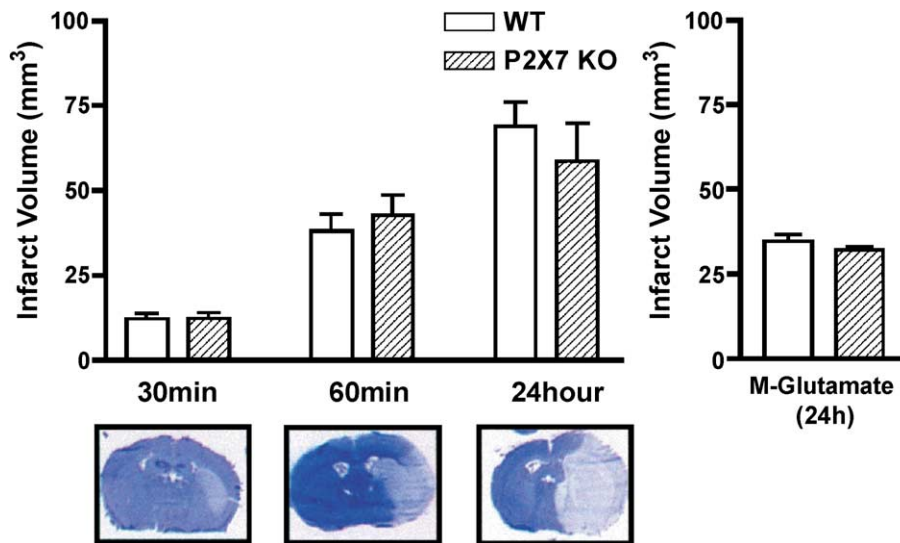


Fig. 3. Absence of the P2X7 receptor does not affect infarct volume following cerebral ischaemia or excitotoxic (methanoglutamate) infusions. Reperfusion (30–60 min) or permanent (24 h) cerebral ischaemia was induced by the intraluminal thread model and excitotoxic lesions by striatal infusion of M-glutamate, in normal mice and those lacking functional P2X7 receptors (P2X7 knockout). Infarct volumes were measured 24 h after surgery on brain sections.

Results generated by the use of knock-out mice must however be treated with caution since compensatory mechanisms may occur. Indeed other P2X receptors may be capable of pore formation by forming heterodimers (e.g., P2X2 and P2X3; [Virginio et al., 1999](#)). However, in vitro studies on primary cultures of macrophages and microglia demonstrated that the effects of ATP on cell death and interleukin-1 β processing and release were completely absent in cells from these P2X7 knockout mice ([Brough et al., 2002](#); [Le Feuvre et al., 2002](#)).

7. Conclusions

It is now generally accepted that high levels of extracellular nucleotides such as ATP may be released under pathological conditions such as inflammation, trauma and stress. Signalling via P2X7 receptors may thus allow cells to sense and respond to events occurring in the extracellular environment, modulate the transcription of genes involved in cellular inflammatory responses and thus to regulate cytokine responses in the CNS ([Liu et al., 2000](#)). In addition to its role in cytokine regulation, it appears that extracellular ATP may act with glutamate and modify intracellular Ca^{2+} homeostasis, and thus have a wider role in neurodegenerative processes ([Fig. 4](#)).

Further research on P2X7 receptors and their actions may unveil novel therapeutic targets to combat neurodegenerative conditions. Future in vivo work is particularly important, as there is often a discrepancy between effects observed in vitro and in vivo. Indeed while interleukin-1 has a profound effect on cell death in vivo, neuroprotective actions of interleukin-1 have been described in vitro ([Strijbos and Rothwell, 1995](#)).

In contrast to the primarily pro-inflammatory activities of nucleotides on immune function, breakdown products of extracellular nucleotides, particularly adenosine, have immunomodulatory properties and neuroprotective roles ([Chen et al., 1999](#); [Dunwiddie and Masino, 2001](#)). A balance between ATP/ADP and adenosine may thus regulate cell death in areas of immune attack by sparing adjacent tissues and possibly by switching the mode of cell death between necrosis and apoptosis. Alterations in P2X7 activation, the presentation/regulation of P2X7 receptors in the

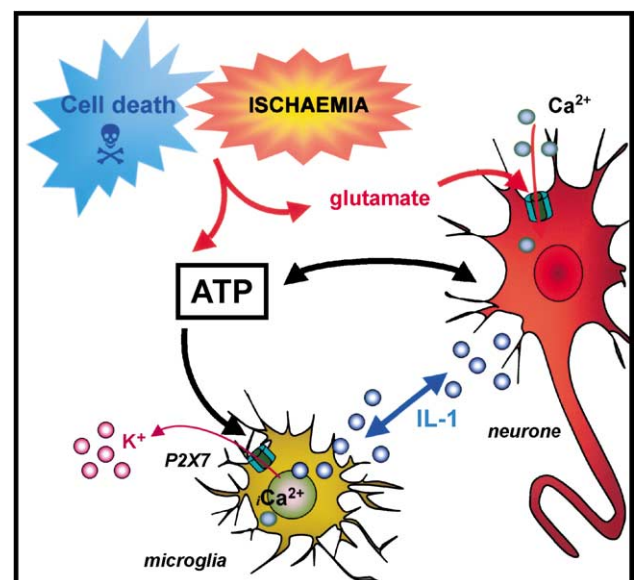


Fig. 4. A general scheme for a role of ATP and P2X7 receptors in neurodegeneration.

plasma membrane (Gu et al., 2000), and its resultant actions could provide interesting therapeutic opportunities.

References

- Allan, S.M., Rothwell, N.J., 2001. Cytokines and acute neurodegeneration. *Nat. Neurosci.* 2, 734–744.
- Andrei, C., Dazzi, C., Lotti, L., Torrisi, M.R., Chimini, G., Rubartelli, A., 1999. The secretory route of the leaderless protein interleukin 1 beta involves exocytosis of endolysosome-related vesicles. *Mol. Biol. Cell* 10, 1463–1475.
- Barone, F.C., Feuerstein, G.Z., 1999. Inflammatory mediators and stroke: new opportunities for novel therapeutics. *J. Cereb. Blood Flow Metab.* 19, 819–834.
- Beigi, R.D., Dubyak, G.R., 2000. Endotoxin activation of macrophages does not induce ATP release and autocrine stimulation of P2 nucleotide receptors. *J. Immunol.* 165, 7189–7198.
- Beigi, R.D., Kobatake, E., Aizawa, M., Dubyak, G.R., 1999. Detection of local ATP release from activated platelets using cell surface-attached firefly luciferase. *Am. J. Physiol.* 276, C267–C278.
- Beuscher, H.U., Gunther, C., Rollinghoff, M., 1990. IL-1 beta is secreted by activated murine macrophages as biologically inactive precursor. *J. Immunol.* 144, 2179–2183.
- Bhat, R.V., DiRocco, R., Marcy, V.R., Flood, D.G., Zhu, Y., Dobrzanski, P., Siman, R., Scott, R., Contreras, P.C., Miller, M., 1996. Increased expression of IL-1 β converting enzyme in hippocampus after ischemia: selective localization in microglia. *J. Neurosci.* 16, 4146–4154.
- Bo, X., Schoepfer, R., Bumstock, G., 2000. Molecular cloning and characterisation of a novel ATP P2X receptor subtype from embryonic chick skeletal muscle. *J. Biol. Chem.* 275, 14401–14407.
- Boutin, H., Le Feuvre, R.A., Horai, R., Asano, M., Iwakura, Y., Rothwell, N.J., 2001. Role of IL-1 α and IL-1 β in ischemic brain damage. *J. Neurosci.* 21, 5528–5534.
- Brough, D., Le Feuvre, R.A., Iwakura, Y., Rothwell, N.J., 2002. Purinergic (P2X7) receptor activation of microglia induces cell death via an interleukin-1 independent mechanism. *Cell. Mol. Neurosci.* 19, 272–280.
- Cerretti, D.P., Kozlosky, C.J., Mosley, B., Nelson, N., Van Ness, K., Greenstreet, T.A., March, C.J., Kronheim, S.R., Druck, T., Cannizzaro, L.A., Huebner, K., Black, R.A., 1992. Molecular cloning of the interleukin-1 β converting enzyme. *Science* 256, 97–102.
- Charriaud-Marlangue, C., Aggoun-Zouaoui, D., Represa, A., Ben-Ari, Y., 1996. Apoptotic features of selective neuronal death in ischemia, epilepsy and gp120 toxicity. *Trends Neurosci.* 19, 109–114.
- Chen, J., Jin, K., Chen, M., Pei, W., Kawaguchi, K., Greenberg, D.A., Simon, R.P., 1997. Early detection of DNA strand breaks in the brain after transient focal ischemia: implications for the role of DNA damage in apoptosis and neuronal cell death. *J. Neurochem.* 69, 232–245.
- Chen, J.F., Huang, Z.H., Ma, J.Y., Zhu, J.M., Moratalla, R., 1999. A2A adenosine receptor deficiency attenuates brain injury induced by transient focal ischemia in mice. *J. Neurosci.* 19, 9200.
- Chiozzi, P., Murgia, M., Falzoni, S., Ferrari, D., Di Virgilio, F., 1996. Role of the purinergic P2z receptor in spontaneous cell death in J774 macrophage cultures. *Biochem. Biophys. Res. Commun.* 218, 176–181.
- Choi, D.W., 1994. Glutamate receptors and the induction of excitotoxic neuronal death. *Prog. Brain Res.* 100, 47–51.
- Ciesynski, J.A., 1999. Calcium ionophore may augment IL-1 secretion by LPS in chick macrophages. *Poult. Sci.* 78, 70–74.
- Collo, G., Neidhart, S., Kawashima, E., Kosco-Vilbois, M., North, R.A., Buell, G., 1997. Tissue distribution of the P2X7 receptor. *Neuropharmacology* 36, 1277–1283.
- Davies, C.A., Loddick, S.A., Toulmond, S., Stroemer, R.P., Hunt, J., Rothwell, N.J., 1999. The progression and topographic distribution of interleukin-1 β expression after permanent middle cerebral artery occlusion in the rat. *J. Cereb. Blood Flow Metab.* 19, 87–98.
- del Zoppo, G., Ginn, J., Hallenbeck, J.M., Iadecola, C., Wang, X., Feuerstein, G.Z., 2000. Inflammation and stroke: putative role for cytokines, adhesion molecules and iNOS in the brain response to ischemia. *Brain Pathol.* 10, 95–112.
- Denlinger, L.C., Fiset, P.L., Sommer, J.A., Watters, J.J., Prabhu, U., Dubyak, G.R., Proctor, R.A., Bertics, P.J., 2001. The nucleotide receptor P2X7 contains multiple protein- and lipid-interaction motifs including a potential binding site for bacterial lipopolysaccharide. *J. Immunol.* 167, 1871–1876.
- Deuchars, S.A., Atkinson, L., Brooke, R.E., Musa, H., Milligan, C.J., Battan, T.F., Buckley, N.J., Parson, S.H., Deuchars, J., 2001. Neuronal P2X7 receptors are targeted to presynaptic terminals in the central and peripheral nervous system. *J. Neurosci.* 21, 7143–7152.
- Di Virgilio, F., 1995. The P2z purinoceptor: an intriguing role in immunity, inflammation and cell death. *Immunol. Today* 16, 524–528.
- Di Virgilio, F., Chiozzi, P., Falzoni, S., Ferrari, D., Sanz, J.M., Venketaraman, V., Baricordi, O.R., 1998a. Cytolytic P2X purinoceptors. *Cell Death Differ.* 5, 191–199.
- Di Virgilio, F., Falzoni, S., Mutini, C., Sanz, J.M., Chiozzi, P., 1998b. Purinergic P2X7 receptor: a pivotal role in inflammation and immunomodulation. *Drug Dev. Res.* 45, 207–213.
- Dinarello, C.A., 1991. Interleukin-1 and interleukin-1 antagonism. *Blood* 77, 1627–1652.
- Dinarello, C.A., 1998. Interleukin-1, interleukin-1 receptors and interleukin-1 receptor antagonist. *Int. Rev. Immunol.* 16, 457–499.
- Dinarello, C.A., Krueger, J.M., 1986. Induction of interleukin 1 by synthetic and naturally occurring muramyl peptides. *Fed. Proc.* 45, 2545–2548.
- Doble, A., 1999. The role of excitotoxicity in neurodegenerative disease: implications for therapy. *Pharmacol. Ther.* 81, 163–221.
- Dubyak, G.R., el-Moatassim, C., 1993. Signal transduction via P2-purinergic receptors for extracellular ATP and other nucleotides. *Am. J. Physiol.* 265, C577–C606.
- Dunwiddie, T.V., Masino, S.A., 2001. The role and regulation of adenosine in the central nervous system. *Annu. Rev. Neurosci.* 24, 31–55.
- Fantuzzi, G., Dinarello, C.A., 1999. Interleukin-18 and interleukin-1 β : two cytokine substrates for ICE (caspase-1). *J. Clin. Immunol.* 19, 1–11.
- Ferrari, D., Chiozzi, P., Falzoni, S., Dal Susino, M., Collo, G., Buell, G., Di Virgilio, F., 1997a. ATP-mediated cytotoxicity in microglial cells. *Neuropharmacology* 36, 1295–1301.
- Ferrari, D., Chiozzi, P., Falzoni, S., Dal Susino, M., Melchiorri, L., Baricordi, O.R., Di Virgilio, F., 1997b. Extracellular ATP triggers IL-1 β release by activating the purinergic P2Z receptor of human macrophages. *J. Immunol.* 159, 1451–1458.
- Ferrari, D., Wesselborg, S., Bauer, M.K.A., Schulze-Osthoff, K., 1997c. Extracellular ATP activates transcription factor NF- κ B through the P2Z purinoceptor by selectively targeting NF- κ B p65 (RelA). *J. Cell Biol.* 139, 1635–1643.
- Ferrari, D., Los, M., Bauer, M.K., Vandenabeele, P., Wesselborg, S., Schulze-Osthoff, K., 1999a. P2Z purinoceptor ligation induces activation of caspases with distinct roles in apoptotic and necrotic alterations of cell death. *FEBS Lett.* 447, 71–75.
- Ferrari, D., Stroth, C., Schulze-Osthoff, K., 1999b. P2X7/P2Z purinoceptor-mediated activation of transcription factor NFAT in microglial cells. *J. Biol. Chem.* 274, 13205–13210.
- Ferrari, D., La Sala, A., Chiozzi, P., Morelli, A., Falzoni, S., Girolomoni, G., Idzko, M., Dichmann, S., Norgauer, J., Di Virgilio, F., 2000. The P2 purinergic receptors of human dendritic cells: identification and coupling to cytokine release. *FASEB J.* 14, 2466–2476.
- Feuerstein, G.Z., Wang, X., Barone, F.C., 1998. Inflammatory mediators and brain injury: the role of cytokines and chemokines in stroke and CNS diseases. In: Ginsberg, M.D., Bogousslavsky, J. (Eds.), *Cerebrovascular Diseases*. Blackwell, Cambridge, pp. 507–531.
- Filippini, A., Taffs, R.E., Agui, T., Sitkovsky, M.V., 1990. Ecto-ATPase activity in cytolytic T-lymphocytes. Protection from the cytotoxic effects of extracellular ATP. *J. Biol. Chem.* 265, 334–340.
- Forloni, G., Bugiani, O., Tagliavini, F., Salmons, M., 1996. Apoptosis-

- mediated neurotoxicity induced by beta-amyloid and PrP fragments. *Mol. Chem. Neuropathol.* 28, 163–171.
- Garcia, J.H., Liu, K.F., Relton, J.K., 1995. Interleukin-1 receptor antagonist decreases the number of necrotic neurons in rats with middle cerebral artery occlusion. *Am. J. Pathol.* 147, 1477–1486.
- Griffiths, R.J., Stam, E.J., Downs, J.T., Otterness, I.G., 1995. ATP induces the release of IL-1 from LPS-primed cells in vivo. *J. Immunol.* 154, 2821–2828.
- Gu, Y., Samecki, C., Aldape, R.A., Livingston, D.J., Su, M.S.S., 1995. Cleavage of poly(ADP-ribose) polymerase by interleukin-1 β converting enzyme and its homologs TX and nedd-2. *J. Biol. Chem.* 270, 18715–18718.
- Gu, B.J., Zhang, W.Y., Bendall, L.J., Chessell, I.P., Buell, G.N., Wiley, J.S., 2000. Expression of P2X(7) purinoceptors on human lymphocytes and monocytes: evidence for nonfunctional P2X(7) receptors. *Am. J. Physiol.* 279, C1189–C1197.
- Haddad, G.G., Jiang, C., 1992. O₂ deprivation in the central nervous system: on mechanisms of neuronal response, differential sensitivity and injury. *Prog. Neurobiol.* 40, 277–318.
- Hamon, Y., Luciani, M.-F., Becq, F., Verrier, B., Rubartelli, A., Chimini, G., 1997. Interleukin-1 β secretion is impaired by inhibitors of the ATP binding cassette transporter, ABC1. *Blood* 90, 2911–2915.
- Hannum, C.H., Wilcox, C.J., Arend, W.P., Joslin, F.G., Dripps, D.J., Heimdal, P.L., Armes, L.G., Sommer, A., Eisenberg, S.P., Thompson, R.C., 2002. Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. *Nature* 434, 336–340.
- Harrison, D.C., Davies, R.P., Bond, B.C., Campbell, C.A., James, M.F., Parson, A.A., Philpott, K.L., 2001. Caspase mRNA expression in a rat model of focal cerebral ischemia. *Brain Res. Mol. Brain Res.* 89, 133–146.
- Hogquist, K.A., Nett, M.A., Unanue, E.R., Chaplin, D.D., 1991. Interleukin-1 is processed and released during apoptosis. *Proc. Natl. Acad. Sci. U. S. A.* 88, 8489.
- Humphreys, B., Rice, J., Kertesz, S., Dubyak, G., 2000. SAPK/JNK activation and apoptotic induction by the macrophage P2X7 nucleotide receptor. *J. Biol. Chem.* 275, 26792–26798.
- Iadecola, C., 1997. Bright and dark sides of nitric oxide in ischemic brain injury. *Trends Neurosci.* 20, 132–139.
- Inoue, K., 1998. The functions of ATP receptors in the hippocampus. *Pharmacol. Res.* 38, 323–331.
- Kim, M., Lee, H.S., LaForet, G., McIntyre, C., Martin, E.J., Chang, P., Kim, T.W., Williams, M., Reddy, P.H., Tagle, S., Boyce, F.M., Won, L., Heller, A., Aronin, N., DiFiglia, M., 1999. Mutant Huntingtin expression in clonal striatal cells: dissociation of inclusion formation and neuronal survival by caspase inhibition. *J. Neurosci.* 19, 964–973.
- Kogure, K., Yamasaki, Y., Matsuo, Y., Kato, H., Onodera, H., 1996. Inflammation in the brain after ischemia. *Acta Neurochir.* 66, 40–43.
- Kreutzberg, G.W., 1996. Microglia: a sensor for pathological events in the CNS. *Trends Neurosci.* 19, 312–318.
- Kristian, T., Siesjö, B.K., 1998. Calcium in ischaemic cell death. *Stroke* 29, 705–718.
- Laliberte, R., Perregaux, D., Svensson, L., Pazoles, C.J., Gabel, C.A., 1994. Tenidap modulates cytoplasmic pH and inhibits anion transport in vitro: II. Inhibition of IL-1 beta production from ATP-treated monocytes and macrophages. *J. Immunol.* 153, 2168–2179.
- Laliberte, R.E., Eggler, J., Gabel, C.A., 1999. ATP treatment of human monocytes promotes caspase-1 maturation and externalization. *J. Biol. Chem.* 274, 36944–36951.
- Lawrence, C.B., Allan, S.M., Rothwell, N.J., 1998. Interleukin-1 β and the IL-1 receptor antagonist act in the striatum to modify excitotoxic brain damage in the rat. *Eur. J. Neurosci.* 10, 1188–1195.
- Lazarowski, E.R., Boucher, R.C., Harden, T.K., 2000. Constitutive release of ATP and evidence for major contribution of ecto-nucleotide pyrophosphatase and nucleoside diphosphokinase to extracellular nucleotide concentration. *J. Biol. Chem.* 275, 31061–31068.
- Lee, J.M., Zipfel, G.J., Choi, D.W., 1999. The changing landscape of ischaemic brain injury mechanisms. *Nature* 399, A7–A14.
- Le Feuvre, R.A., Brough, D., Iwakura, Y., Takeda, K., Rothwell, N.J., 2002. Priming of macrophages with LPS potentiates P2X7 mediated cell death via a caspase-1 dependent mechanism, independently of cytokine production. *J. Biol. Chem.* 277, 3210–3218.
- Legos, J.J., Whitmore, R.G., Erhardt, J.A., Parsons, A.A., 2000. Quantitative changes in interleukin proteins following focal stroke in the rat. *Neurosci. Lett.* 282, 189–192.
- Li, P., Allen, S., Banerjee, S., Franklin, L., Herzog, C., Johnston, J., McDowell, J., Paskind, L., Rodman, L., Salfeld, J., 1995. Mice deficient in IL-1 beta converting enzyme are defective in production of mature IL-1 and resistant to endotoxic shock. *Cell* 80, 401–411.
- Limbrick, D.D., Churn, S.B., Sombati, S., Delorenzo, R.J., 1995. Inability to restore resting intracellular calcium levels as an early indicator of delayed neuronal cell death. *Brain Res.* 690, 145–156.
- Lipton, S.A., Rossenberger, P.A., 1994. Excitatory amino acids as a final common pathway for neurologic disorders. *N. Engl. J. Med.* 330, 613–622.
- Liu, X.H., Kwon, D., Schielke, G.P., Yang, G.Y., Silverstein, F.S., Barks, J.D., 1999. Mice deficient in interleukin-1 converting enzyme are resistant to neonatal hypoxic–ischemic brain damage [In Process Citation]. *J. Cereb. Blood Flow Metab.* 19, 1099–1108.
- Liu, J.S.H., John, G.R., Sikora, I.A., Lee, S.C., Brosnan, C.F., 2000. Modulation of interleukin-1 β and tumour necrosis factor α signalling by P2 purinergic receptors in human fetal astrocytes. *J. Neurosci.* 20, 5292–5299.
- Loddick, S.A., Rothwell, N.J., 1996. Neuroprotective effects of human recombinant interleukin-1 receptor antagonist in focal cerebral ischemia in the rat. *J. Cereb. Blood Flow Metab.* 16, 932–940.
- Loddick, S.A., Wong, M.L., Bongiorno, P.B., Gold, P.W., Licinio, J., Rothwell, N.J., 1997. Endogenous interleukin-1 receptor antagonist is neuroprotective. *Biochem. Biophys. Res. Commun.* 234, 211–215.
- Mabuchi, T., Kitagawa, K., Ohtsuki, T., Kuwabara, K., Yagita, Y., Yanagihara, T., Hori, M., Matsumoto, M., 2000. Contribution of microglia/macrophages to expansion of infarction and response of oligodendrocytes after focal cerebral ischaemia in rats. *Stroke* 31, 1735–1743.
- MacKenzie, A., Wilson, H.L., Kiss-Toth, E., Dower, S.K., North, A., Surprenant, A., 2001. Rapid secretion of interleukin-1 β by microvesicle shedding. *Immunity* 8, 825–835.
- Mattson, M.P., 1998. Modification of ion homeostasis by lipid peroxidation: roles in neuronal degeneration and adaptive plasticity. *Trends Neurosci.* 21, 53–57.
- McCulloch, J., Ozyurt, E., Park, C.K., Nehls, D.G., Teasdale, G.M., Graham, D.I., 1993. Glutamate receptor antagonists in experimental focal cerebral ischemia. *Acta Neurochir.* 57, 73–79.
- Mehta, V.B., Hart, J., Wewers, M.D., 2001. ATP stimulated release of IL-1 beta and IL-18 requires priming by LPS and is independent of caspase-1 cleavage. *J. Biol. Chem.* 276, 3820–3826.
- Molloy, A., Laochumroonvorapong, P., Kaplan, G., 1994. Apoptosis, but not necrosis, of infected monocytes is coupled with killing of intracellular bacillus Calmette–Guerin. *J. Exp. Med.* 180, 1499–1509.
- Mutini, C., Falzoni, S., Ferrari, D., Chiozzi, P., Morelli, A., Baricordi, O.R., Collo, G., Ricciardi-Castagnoli, P., Di Virgilio, F., 1999. Mouse dendritic cells express the P2X7 purinergic receptor: characterization and possible participation in antigen presentation. *J. Immunol.* 163, 1958–1965.
- Nakatsuka, T., Gu, J.G., 2001. ATP P2X receptor mediated enhancement of glutamate release and evoked EPSC's in dorsal horn nucleus of the rat spinal cord. *J. Neurosci.* 21, 6522–6531.
- Nicholson, D.W., Thornberry, N.A., 1997. Caspases: killer proteases. *TIBS* 22, 299–306.
- Nicotra, P., 2000. Apoptosis and neurodegeneration: the role of caspases. In: Kriegstein, J., Klumpp, S. (Eds.), *Pharmacology of Cerebral Ischemia*. Medpharm Scientific Publishers, Stuttgart, pp. 3–9.
- Nieber, K., Eschke, D., Brand, A., 1999. Brain hypoxia: effects of ATP and adenosine. *Prog. Brain Res.* 120, 287–300.
- O'Neill, L.A., Dinarello, C.A., 2000. The IL-1 receptor/toll like receptor superfamily: crucial receptors for inflammation and host defence. *Immunol. Today* 21, 206–209.

- Paschen, W., Doutheil, J., 1999. Disturbances of the functioning of endoplasmic reticulum: a key mechanism underlying neuronal cell injury? *J. Cereb. Blood Flow Metab.* 19, 1–18.
- Pearson, V.L., Rothwell, N.J., Toulmond, S., 1999. Excitotoxic brain damage in the rat induces interleukin-1 beta protein in microglia and astrocytes: correlation with the progression of cell death. *Glia* 25, 311–323.
- Perregaux, D., Gabel, C.A., 1994. Interleukin-1 beta maturation and release in response to ATP and nigericin. Evidence that potassium depletion mediated by these agents is a necessary and common feature of their activity. *J. Biol. Chem.* 269, 15195–15203.
- Perregaux, D.G., Gabel, C.A., 1998. Post-translational processing of murine IL-1: evidence that ATP-induced release of IL-1 alpha and IL-1 beta occurs via a similar mechanism. *J. Immunol.* 160, 2469–2477.
- Perregaux, D., Barberia, J., Lanzetti, A.J., Geoghegan, K.F., Carty, T.J., Gabel, C.A., 1992. IL-1 beta maturation: evidence that mature cytokine formation can be induced specifically by nigericin. *J. Immunol.* 149, 1294–1303.
- Perregaux, D.G., Svensson, L., Gabel, C.A., 1996. Tenidap and other anion transport inhibitors disrupt cytolytic T lymphocyte-mediated IL-1 beta post-translational processing. *J. Immunol.* 157, 57–64.
- Raivich, G., Bohatschek, M., Kloss, C.U.A., Werner, A., Jones, L.L., Kreutzberg, G.W., 1999. Neuroglial activation in the injured brain: graded response, molecular mechanisms and cues to physiological function. *Brain Res. Rev.* 30, 77–105.
- Ralevic, V., Burnstock, G., 1998. Receptors for purines and pyrimidines. *Pharmacol. Rev.* 50, 413–492.
- Rassendren, F., Buell, G., Newbolt, A., North, R.A., Surprenant, A., 1997. Identification of amino acid residues contributing to the pore of a P2X receptor. *EMBO J.* 16, 3446–3454.
- Relton, J.K., Rothwell, N.J., 1992. Interleukin-1 receptor antagonist inhibits ischaemic and excitotoxic neuronal damage in the rat. *Brain Res. Bull.* 29, 43–46.
- Roux-Lombard, P., 1998. The interleukin-1 family. *Eur. Cytokine Network* 9, 565–576.
- Sanz, J.M., Di Virgilio, F., 2000. Kinetics and mechanism of ATP-dependent IL-1 β release from microglial cells. *J. Immunol.* 164, 4893–4898.
- Sanz, J.M., Chiozzi, P., Di Virgilio, F., 1998. Tenidap enhances P2Z/P2X7 receptor signalling in macrophages. *Eur. J. Pharmacol.* 355, 235–244.
- Sattler, R., Xiang, Z., Lu, W.Y., MacDonald, J.F., Tymianski, M., 1999. Specific coupling of NMDA receptor activation to nitric oxide neurotoxicity by PSD-95 protein. *Science* 284, 1845–1848.
- Saudou, F., Finkbeiner, S., Devys, D., Greenberg, M.E., 1998. Huntingtin acts in the nucleus to induce apoptosis but death does not correlate with the formation of intranuclear inclusions. *Cell* 95, 55–66.
- Savinainen, A., Garcia, E.P., Dorow, D., Marshall, J., Liu, Y.F., 2001. Kainate receptor activation induces mixed lineage kinase-mediated cellular signaling cascades via post-synaptic density protein 95. *J. Biol. Chem.* 276, 11382–11386.
- Schneider, A., Martin-Villalba, F., Weih, F., Vogel, J., Wirth, T., Schwanner, M., 2000. NF- κ B is activated and promotes cell death in focal cerebral ischaemia. In: Kriegstein, J., Klumpp, S. (Eds.), *Pharmacology of Cerebral Ischemia*. Medpharm Scientific Publishers, Stuttgart, pp. 133–136.
- Schulze-Lohoff, E., Hugo, C., Rost, S., Arnold, S., Gruber, A., Brune, B., Sterzel, R.B., 1998. Extracellular ATP causes apoptosis and necrosis of cultured mesangial cells via P2Z/P2X7 receptors. *Am. J. Physiol.* 275, F962–F971.
- Siesjö, B.K., 1981. Cell damage in the brain: a speculative synthesis. *J. Cereb. Blood Flow Metab.* 1, 155–185.
- Sikora, A., Liu, J., Brosnan, C., Buell, G., Chessel, I., Bloom, B.R., 1999. Cutting edge: purinergic signaling regulates radical-mediated bacterial killing mechanisms in macrophages through a P2X7-independent mechanism. *J. Immunol.* 163, 558–561.
- Sims, J.E., Gayle, M.A., Slack, J.L., Alderson, M.R., Bird, T.A., Giri, J.G., Colotta, F., Re, F., Mantovani, A., Shanebeck, K., 1993. Interleukin-1 signalling occurs exclusively via the type 1 receptor. *Proc. Natl. Acad. Sci.* 90, 6155–6159.
- Singer, I.I., Scott, S., Chin, J., Bayne, E.K., Limjoco, G., Weidner, J., Miller, D.K., Chapman, K., Kostura, M.J., 1995. The interleukin-1 beta-converting enzyme (ICE) is localized on the external cell surface membranes and in the cytoplasmic ground substance of human monocytes by immuno-electron microscopy. *J. Exp. Med.* 182, 1447–1459.
- Smith, D.E., Renshaw, B.R., Kechem, R.R., Kubin, M., Garka, K.E., Sims, J.E., 2000. Four new members expand the interleukin-1 superfamily. *J. Biol. Chem.* 275, 1169–1175.
- Solle, M., Labasi, J., Perregaux, D.G., Stam, E., Petrushova, N., Koller, B.H., Griffiths, R.J., Gabel, C.A., 2001. Altered cytokine production in mice lacking P2X7 receptors. *J. Biol. Chem.* 276, 125–132.
- Sperlágh, B., Vizi, E.S., 1996. Neuronal synthesis, storage and release of ATP. *Semin. Neurosci.* 8, 175–186.
- Stevenson, F.T., Bursten, S.L., Fanton, C., Locksley, R.M., Lovett, D.H., 1993. The 31-kDa precursor of interleukin 1 alpha is myristoylated on specific lysines within the 16 kDa N-terminal propeptide. *Proc. Natl. Acad. Sci.* 90, 7245–7249.
- Stoll, G., Jander, S., Schroeter, M., 1998. Inflammation and glial responses in ischemic brain lesions. *Prog. Neurobiol.* 56, 149–171.
- Strijbos, P.J.L.M., Rothwell, N.J., 1995. Interleukin-1 β attenuates excitatory amino acid-induced neurodegeneration in vitro: involvement of nerve growth factor. *J. Neurosci.* 15, 3468–3474.
- Stroemer, R.P., Rothwell, N.J., 1998. Exacerbation of ischemic brain damage by localized striatal injection of interleukin-1 β in the rat. *J. Cereb. Blood Flow Metab.* 18, 833–839.
- Surprenant, A., Rassendren, F., Kawashima, E., North, R.A., Buell, G., 1996. The cytolytic P_{2Z} receptor for extracellular ATP identified as a P2X receptor (P2X7). *Science* 272, 735–738.
- Thornberry, N.A., Bull, H.G., Calaycay, J.R., Chapman, K.T., Howard, A.D., Kostura, M.J., Miller, D.K., Molineaux, S.M., Weidner, J.R., Aunins, J., 1992. A novel heterodimeric cysteine protease is required for interleukin-1 beta processing in monocytes. *Nature* 356, 768–774.
- Touzani, O., Boutin, H., Chequet, J., Rothwell, N.J., 1999. Potential mechanisms of interleukin-1 involvement in cerebral ischemia. *J. Neuroimmunol.* 100, 203–215.
- Touzani, O., Boutin, H., Le Feuvre, R.A., Parker, L., Miller, A., Luheshi, G., Rothwell, N.J., 2002. Interleukin-1 influences ischemic brain damage in the mouse independently of the interleukin-1 type 1 receptor. *J. Neurosci.* 22, 38–43.
- Vey, E., Dayer, J.M., Burger, D., 1997. Direct contact with stimulated T cells induces the expression of IL-1 beta and IL-1 receptor antagonist in human monocytes. Involvement of serine/threonine phosphatases in differential regulation. *Cytokine* 9, 480–487.
- Virginio, C., MacKenzie, A., Rassendren, F.A., North, R.A., Surprenant, A., 1999. Pore dilation of neuronal P2X receptor channels. *Nat. Neurosci.* 2, 315–321.
- Valev, I., Reske, K., Palmer, M., Valeva, A., Bhakdi, S., 1995. Potassium-inhibited processing of IL-1 beta in human monocytes. *EMBO J.* 14, 1607–1614.
- Valev, I., Klein, J., Husmann, M., Valeva, A., Strauch, S., Wirtz, H., Weichel, O., Bhakdi, S., 2000. Potassium regulates IL-1 beta processing via calcium-independent phospholipase A2. *J. Immunol.* 164, 5120–5124.
- Watanabe, N., Kawaguchi, M., Kobayashi, Y., 1998. Activation of interleukin-1 beta-converting enzyme by nigericin is independent of apoptosis. *Cytokine* 10, 645–653.
- West, M.A., Clair, L., Bellingham, J., 1996. Role of calcium in lipopolysaccharide-stimulated tumour necrosis factor and interleukin-1 signal transduction in naive and endotoxin-tolerant murine macrophages. *J. Trauma* 41, 647–652.
- Wilson, K.P., Black, J.-A.F., Thomson, J.A., Kim, E.E., Griffith, J.P., Navia, M.A., Murcko, M.A., Chambers, S.P., Aldape, R.A., Raybuck, S.A., Livingston, D.J., 1994. Structure and mechanism of interleukin-1 β converting enzyme. *Nature* 370, 270–275.
- Yamasaki, Y., Matsuura, N., Shozuhara, H., Onodera, H., Itoyama, Y., Kogure, K., 1995. Interleukin-1 as a pathogenic mediator of ischemic brain damage in rats. *Stroke* 26, 676–681.